

1 **Supplementary figure legends**

2 **Supplementary Figure 1. Modulation of axonal retrograde transport is**

3 **detected by the accumulation assay. (A-D)** Treatment of ES cell-derived

4 motor neurons with 1 mM EHNA causes a significant decrease in the

5 accumulation of H_CT (A) and α-p75^{NTR} (C) in the cell body. ES cell-derived

6 motor neurons were incubated with AlexaFluor 555-conjugated H_CT or α-

7 p75^{NTR} and either DMSO or 1 mM EHNA for 2 h. Cells were then acid-

8 washed, fixed, permeabilised and stained for α-p75^{NTR}. The amount of

9 intracellular H_CT (B) and α-p75^{NTR} (D) was quantified as the mean staining

10 intensity per pixel in the cell body. (E-F) Defective transport in SOD1^{G93A}

11 motor neurons is observed as a decrease in accumulation of H_CT in the cell

12 body. Primary motor neurons from E13 SOD1^{G93A} embryos were incubated

13 with AlexaFluor 555-conjugated H_CT for 2 h. Cells were then acid-washed and

14 fixed. The amount of intracellular H_CT (F) was quantified as the mean staining

15 intensity per pixel in the cell body. Results are expressed as a percentage of

16 the control (A-D) or wild type (E-F) (n ≥ 25 cell bodies per condition; n = 3

17 independent experiments). *** p< 0.001, **** p< 0.0001 (unpaired Student's *t*

18 test). Scale bars, 10 μm.

19

20 **Supplementary Figure 2. Compound A1 accelerates axonal transport in**

21 **SOD1^{G93A} but not in wild type motor neurons. (A)** The results of a small

22 molecule chemical screen are shown as an XY plot of the normalized mean

23 staining intensity for α-p75^{NTR} versus AlexaFluor 555-conjugated H_CT. α-p75^{NTR}

24 was detected as described in Figure 2. The red dot highlights the lack of effect of

25 a reported JNK inhibitor in this assay. This result is in line with data shown in

Figure 5. The yellow rectangle corresponds to three times the standard deviation of the dataset. The negative control (EHNA) is shown in blue ($n \geq 25$ cell bodies analysed per condition; $n = 3$ repeats). **(B)** Speed profiles of H_CT carriers in wild type motor neurons treated with DMSO (black squares) or 2 μ M A1 (light blue triangles). Comparison of the curves reveals that compound A1 has no effect on the speed of axonal retrograde transport in wild type neurons (wild type: 94 carriers, 13 axons; wild type + 2 μ M A1: 108 carriers, 11 axons; 4 independent experiments).

Supplementary Figure 3. Target validation of the small molecule screen.

(A) Summary of the results of the small molecule chemical screen plotted as the normalized mean staining intensity values for α -p75^{NTR} versus H_CT. An additional fourteen p38 MAPK inhibitors were screened for their effect on the accumulation of H_CT and α -p75^{NTR}. Active compounds were defined as those able to increase the accumulation of α -p75^{NTR} and H_CT by at least three times the standard deviation of the entire dataset (represented by the yellow rectangle); $n \geq 25$ cell bodies per condition, $n = 3$ repeats per conditions. The active compounds are shown in red, whilst the negative control (EHNA) is shown in blue. **(B-E)** Speed profiles of AlexaFluor 555-conjugated H_CT carriers in SOD1^{G93A} motor neurons treated with 2 μ M of each of the active compounds (wild type: 74 carriers, 8 axons; SOD1^{G93A}: 119 carriers, 7 axons; SOD1^{G93A} + 2 μ M A3: 120 carriers, 7 axons (B); SOD1^{G93A} + 2 μ M C3: 117 carriers, 8 axons (C); SOD1^{G93A} + 2 μ M D3: 166 carriers, 9 axons (D); SOD1^{G93A} + 2 μ M H2: 94 carriers, 7 axons; 3 independent experiments (E)).

Supplementary Figure 4. p38 MAPK is activated in both embryonic motor neurons and adult SOD1^{G93A} spinal cord. Western blot showing levels of phospho-p38 MAPK in spinal cord lysates from wild type, SOD1^{G93A} and SOD1^{WT} overexpressing mice (*top*). Active p38 MAPK was detected with a phosphospecific pT180/pY182 α -p38 MAPK antibody, whilst the total content of p38 MAPK was assessed by a pan α -p38 MAPK antibody. The ratios of p-p38 MAPK / total p38 MAPK α signals are shown in the lower part of the panel. The ratios of mutant and wild type hSOD1 at different ages were been normalised to the ratio of age matched wild type littermates (taken as 1). p38 MAPK shows increased activation in pre-symptomatic (36 d) and early symptomatic (73 d) SOD1^{G93A} mice. Spinal cords from SOD1^{WT} overexpressing mice show no change in p38 MAPK activation compared to wild type controls (n = 1 animal per condition).

Supplementary Figure 5. Lentiviral-mediated transduction of primary motor neurons. (A) Western blot showing lentivirus-mediated knockdown of p38 MAPK α in primary motor neurons (n = 1 experiment). (B) Quantification of western blots shows a reduction in p38 MAPK α levels with p38 MAPK α -targeted shRNA, whilst scrambled shRNA has no effect. (C) Example of western blot showing the lentivirus-mediated knockdown of p38 MAPK δ (*left*) and its quantification showing dose-dependent reduction of p38 MAPK δ expression (*right*). (D) Speed profiles of H_CT carriers in SOD1^{G93A} motor neurons transduced with scrambled shRNA (scram; red diamonds) and SOD1^{G93A} motor neurons transduced with p38 MAPK δ shRNA (brown triangles). Comparison of the curves reveals that knockdown of p38 MAPK δ has no effect on axonal transport speeds

(SOD1^{G93A} + scrambled shRNA: 46 carriers, 10 axons; SOD1^{G93A} + p38 MAPK δ shRNA: 101 carriers, 12 axons; 3 independent experiments).

Supplementary Figure 6. SB-239063 affects neurofilament phosphorylation in primary motor neurons. (A) *Top panel:* Effect of 2 μ M SB-239063 on the phosphorylation of neurofilament heavy chain detected by the SMI34 antibody in motor neurons isolated from wild type and SOD1^{G93A} mice. Actin was used as loading control (n = 3 independent experiments). *Lower panel:* western blot quantification revealed a reduction in the SMI34 signal in extracts from SOD1^{G93A} cultures after 24 h of treatment (n = 3 independent experiments). Data shown as mean \pm SEM. ** p< 0.01 (two-way ANOVA followed by Tukey's comparison test). (B) Wild type mice were administered either 30 mg/kg or 100 mg/kg SB-239063 i.p. The free concentration in the brain, spinal cord and muscle was determined after 1 h. ^ based on fraction unbound in rat brain. ^^ based on fraction unbound in rat blood. (C) Predicted brain free concentration profiles upon administration of 30 mg/kg and 100 mg/kg of SB-239063 via i.p. at 0 and 8 h.

Supplementary Figure 7. Chronic treatment with SB-239063 does not restore axonal transport deficits in 70 day old SOD1^{G93A} mice and has severe side effects. (A) Muscle endplate occupancy in lumbricals muscles of the distal hindlimb. Examples show fully innervated endplates, with neurofilament heavy chain and synaptic vesicle protein 2 (NF/SV2, in green) labelling covering the endplate region (labelled by α -bungarotoxin (BTx) in red), indicating fully innervated endplates (arrows). Some endplates only

display partial neurofilament coverage, indicating these endplates are partially innervated (arrowheads), whereas in some cases endplates do not show any labelling for neurofilament/SV2, an indication for denervated endplates (double arrows). Scale bars, 20 μ m. **(B)** Muscle endplate occupancy in lumbricals muscles of the distal hindlimb was assessed in wild type, vehicle-treated and SB-239063 treated SOD1^{G93A} mice at 90 d (*left*) and 120 d (*right*) of age. Endplate occupancy was measured as the percentage of the total number of endplates assessed (n = minimum of 3 mice in each experimental group). The level of denervation was compared between each experimental group using a 2 way ANOVA with Tukeys multiple comparisons test. At 90 d there was significant denervation in SOD1^{G93A} mice compared to controls (p=0.0264), which was also present at a later stage (120 d; p=0.0044). Treatment with SB-239063 did not result in a significant protection from denervation compared to vehicle-treated SOD1^{G93A} mice (p=0.9998 at 90 d; p=0.8934 at 120 d). **(C)** Timeline of treatment of SOD1^{G93A} mice with SB-239063. Mice were injected i.p. with 10 mg/kg SB-239063 twice daily until the day of experiment. **(D)** The axonal transport of AlexaFluor 555-conjugated H_CT cargoes was measured in live anaesthetized mice at 120 d (wild type: 128 carriers, n (animals) = 5; SOD1^{G93A} + vehicle: 110 carriers, n = 4; SOD1^{G93A} + 100 mg/kg SB-239063: 139 carriers, n = 5; data shown as mean \pm SEM). **(E)** Timeline of treatment of SOD1^{G93A} mice with SB-239063. Mice were injected i.p. with 10 mg/kg SB-239063 twice daily until the day of experiments. **(F)** Axonal transport in SOD1^{G93A} mice at 70 d is not improved by long-term treatment with SB-239063 starting at 50 d (wild type: 187 carriers, n (animals) = 5; SOD1^{G93A} + vehicle: 103 carriers, n = 3; SOD1^{G93A} + 10 mg/kg SB-239063:

126 136 carriers, n = 4; data shown as mean \pm SEM). **(G)** SOD1^{G93A} mice treated
127 chronically, twice daily, with SB-239063 go through an initial phase of weight
128 loss, compared to vehicle treated SOD1^{G93A} mice (wild type: n (animals) = 8;
129 SOD1^{G93A} + vehicle: n = 9; SOD1^{G93A} + 10 mg/kg SB-239063: n = 7; data shown
130 as mean \pm SEM). **(H)** Chronic treatment with either vehicle or SB-239063 leads
131 to severe spleno- and hepato-megaly in SOD1^{G93A} mice. Grid size: 10 mm.
132

133 **Supplementary Table 1. Library of kinase inhibitors used for the primary**
134 **screen.** Selected active compounds are highlighted in grey.

Reference code	Compound name	Primary Target
2A1	GW693481X	TGFBR
2A2	GW572738X	JNK
3A1	GSK200398A	EGFR/ErbB2
3A2	GW651576X	EGFR/ErbB2
A1	SB-239272	p38 MAPK
A2	GSK248233A	RHO
A3	GW296115X	PDGFR
A4	GW589961A	TIE2/VEGFR2
A5	GW806742X	VEGFR
A6	GSK586581A	IKK
A7	GW410563A	VEGFR
B1	GW801372X	GSK3
B2	SB-772077-B	RHO
B3	SB-400868-A	TGFBR
B4	SB-220025-A	p38 MAPK
B5	GI261520A	EGFR/ErbB2
B6	GW824645A	CDK2
B7	GSK1023156A	PLK
C1	GW679410X	TGFBR
C2	SB-675259-M	GSK3
C3	GW440139A	RET
C4	GW820759X	p38 MAPK
C5	GW642125X	TIE2/VEGFR2
C6	GW275616X	TRKA
C7	GW406731X	RAF
D1	GW549390X	VEGFR
D2	GW561436X	p38 MAPK
D3	GW768505A	TIE2/VEGFR2
D4	GW856804X	TIE2/VEGFR2
D5	GW461104A	EGFR/ErbB2
D6	GSK180736A	RHO
D7	GW559768X	RET
E1	GW575808A	LCK
E2	GSK319347A	IKK
E3	GW743024X	p38 MAPK
E5	GSK1000163A	AKT
E6	GSK237700A	PLK
F1	GW632580X	CSF
F2	GW837331X	PLK
F3	SB-358518	GSK3
F4	GW275944X	CDK2
F5	GSK554170A	AKT
F6	GW513184X	GSK3

G1	GSK317354A	RHO
G2	GW284372X	EGFR/ErbB2
G3	SB-437013	TIE2
G4	GW305074X	RAF
G5	SB-737198	MSK
G6	GSK1173862A	IGF-1R
H1	GW819230X	GSK3
H2	SB-744941	MSK
H3	SB-725317	GSK3
H4	GSK182497A	EGFR/ErbB2
H5	GR105659X	TRKA
H6	GSK953913A	IKK

135

136 **Supplementary Table 2. Library of kinase inhibitors used for the**
 137 **validation (secondary) screen.** Selected active compounds are highlighted
 138 in grey.

Reference code	Compound name	Primary Target
A1	SB-226879	p38 MAPK
A2	GW775608X	p38 MAPK
A3	GW796921X	p38 MAPK
B1	SB-221466	p38 MAPK
B2	GW581744X	p38 MAPK
C2	GW607117X	p38 MAPK
C3	SB-223133	p38 MAPK
D1	GW618013A	p38 MAPK
D2	SB-242719	p38 MAPK
D3	GW434756X	p38 MAPK
E2	GW734508X	p38 MAPK
G1	GW769076X	p38 MAPK
H1	GW569293E	p38 MAPK
H2	GW782907X	p38 MAPK

139

140 **Supplementary Table 3. Active p38 MAPK α compounds used *in vitro***
 141 **and/or *in vivo* in this study.**

Compound name	Primary Target	ChEMBL[#]	References
SB-239063	p38 MAPK α	97162	(8, 9)
SB-203580	p38 MAPK α	10	(8)
SB-239272 (A1)	p38 MAPK α	275798	(8)
SB-223133	p38 MAPK α	1888586	(8)
GW796921X	p38 MAPK α	517666	(10)
GW434756X	p38 MAPK α	485745	(11)
GW782907X	p38 MAPK α	477978	(10)

142 [#]<https://www.ebi.ac.uk/chembl/>

Figure S1

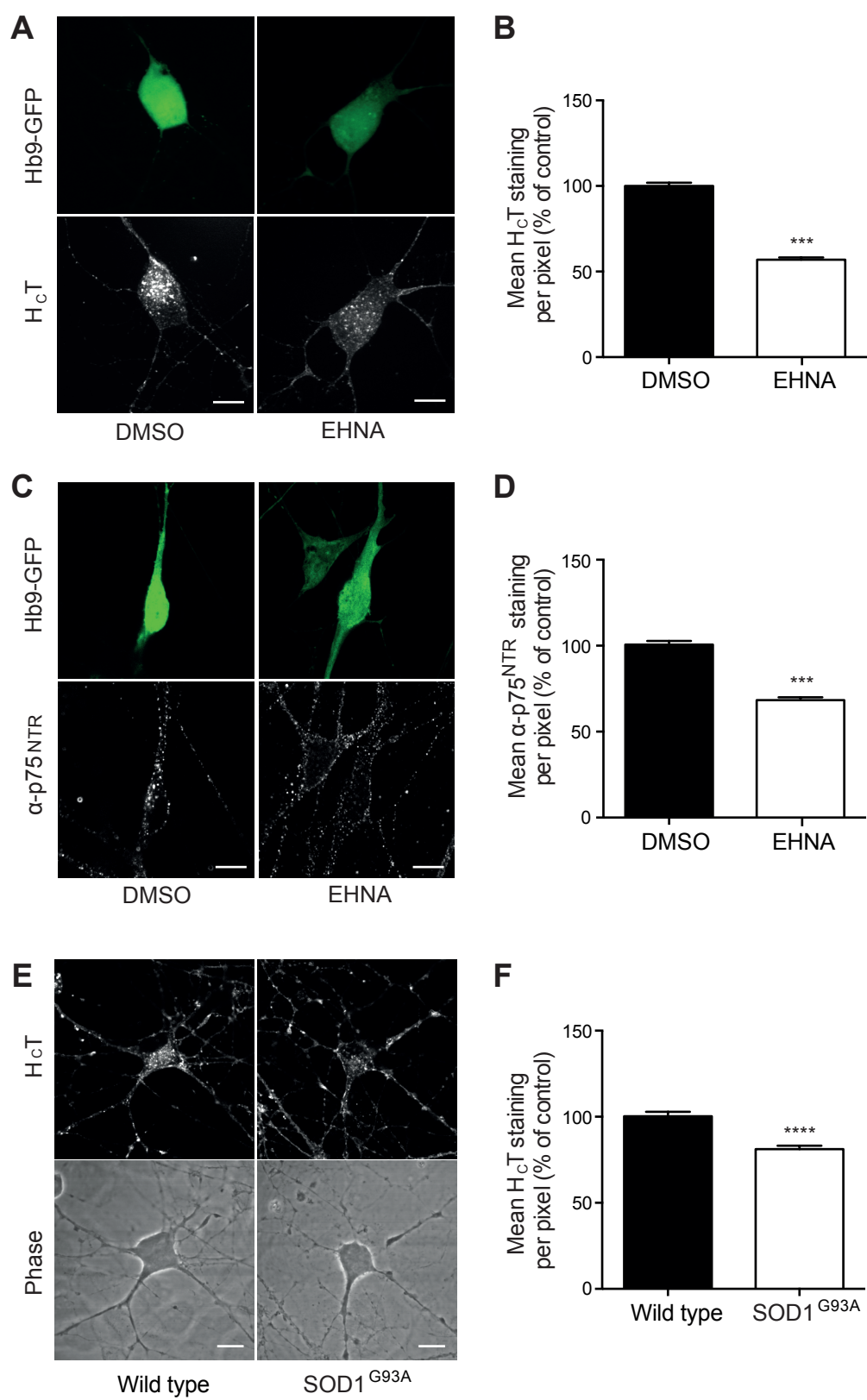


Figure S2

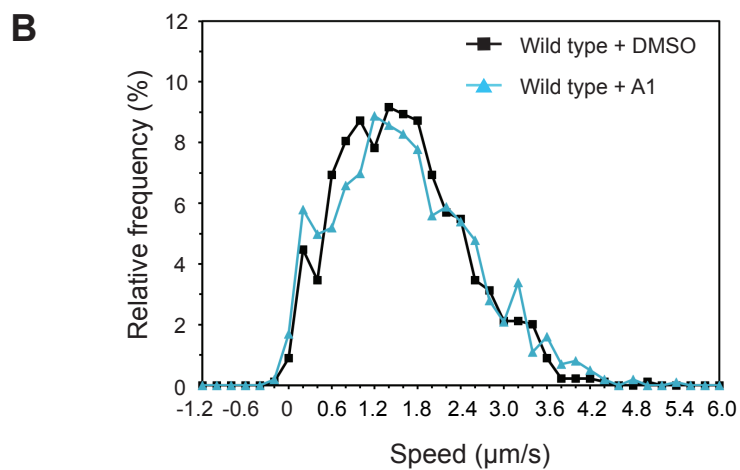
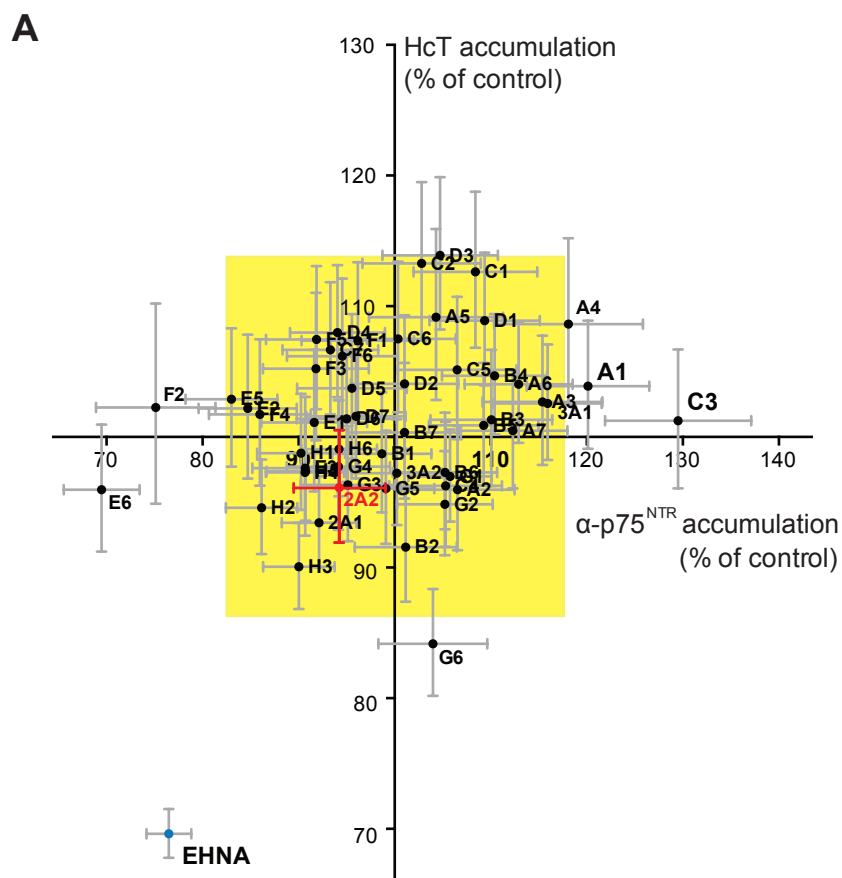
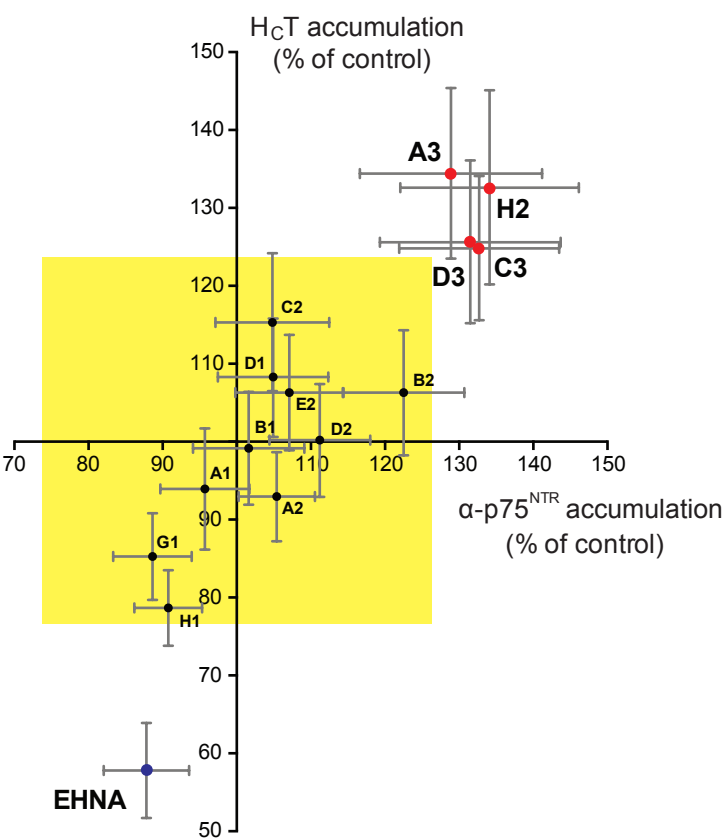
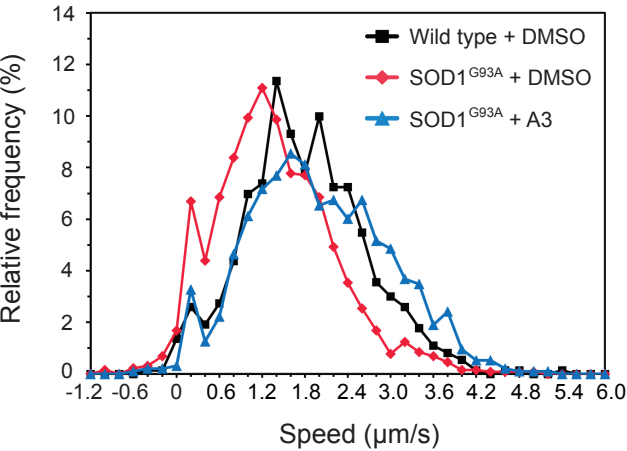


Figure S3

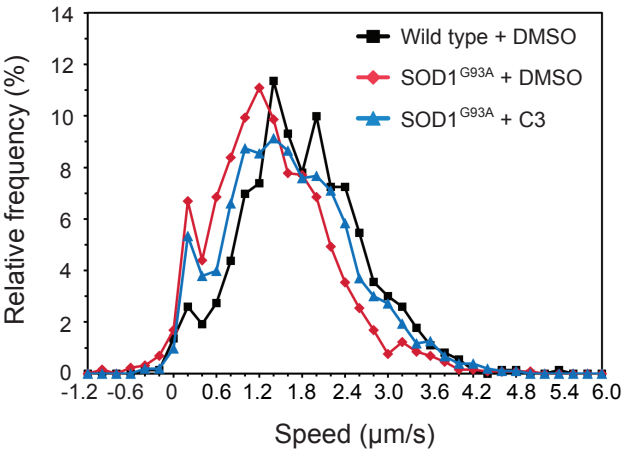
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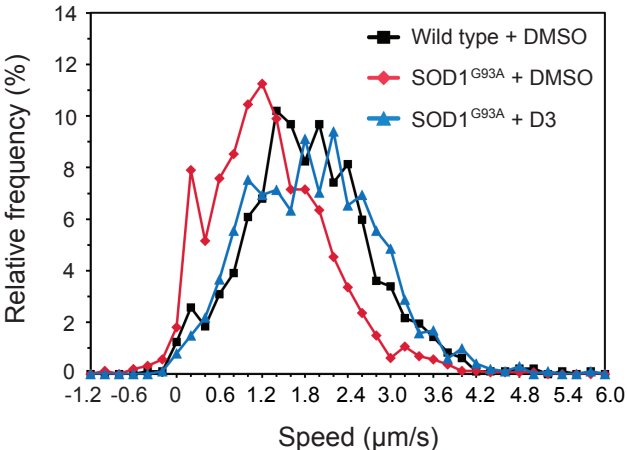
B



C



D



E

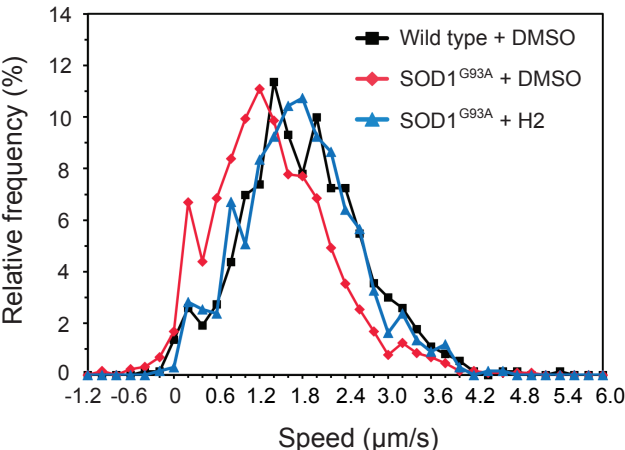


Figure S4

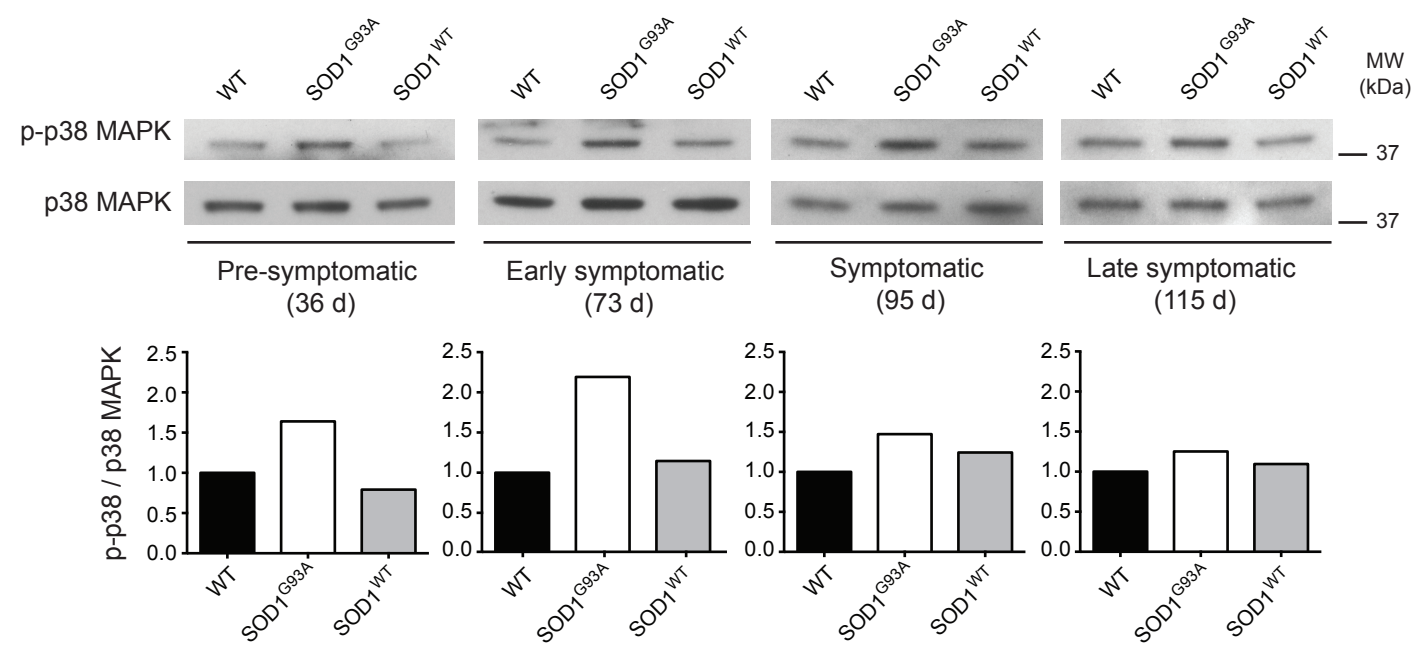


Figure S5

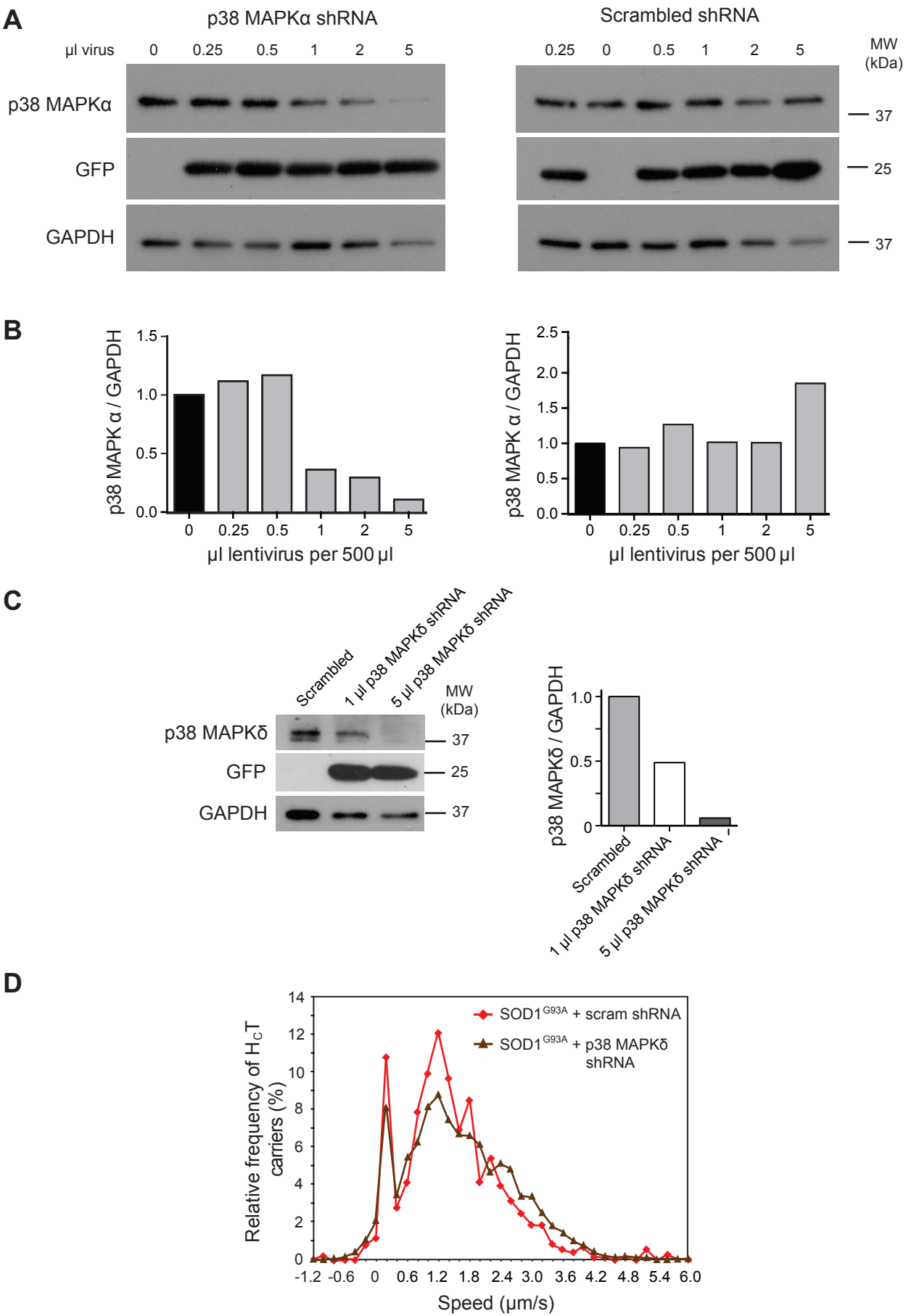
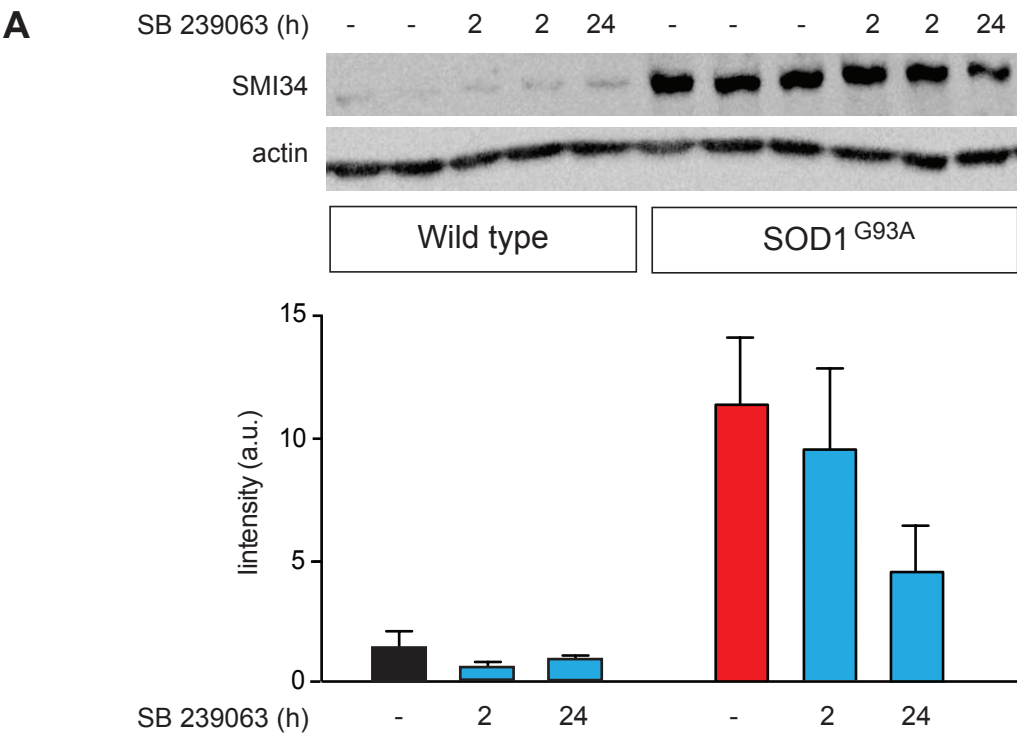


Figure S6



B

Dose (mg/kg)	Time (h)	Brain (μM) ^	Spinal Cord (μM) ^	Muscle (μM) ^^
30	1	0.464	0.405	1.182
30	1	0.555	0.546	1.274
100	1	1.784	1.82	3.241
100	1	1.337	1.379	3.476

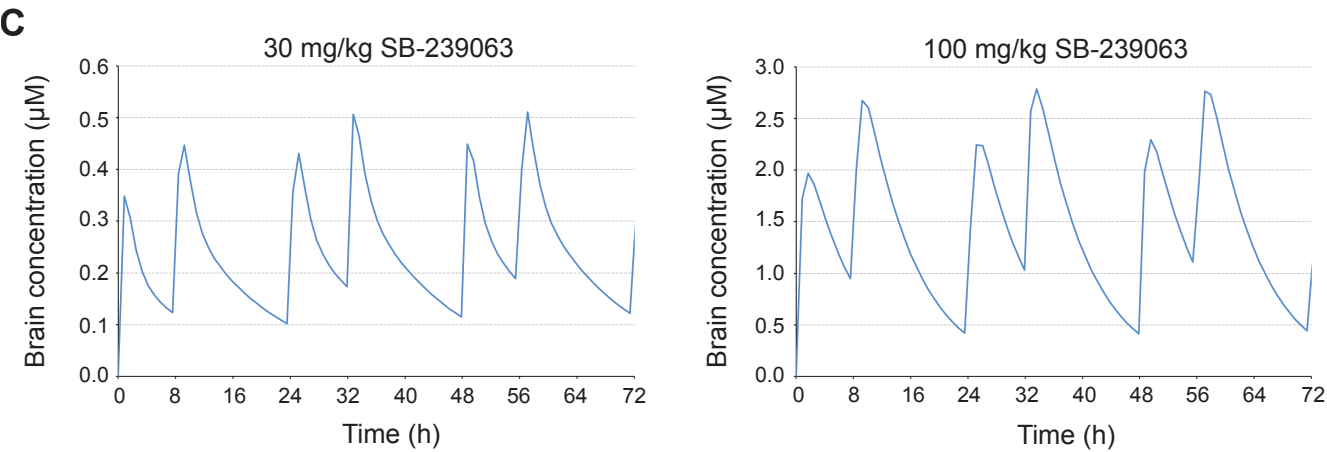


Figure S7

